

Emulating the Logic of Monoterpenoid Alkaloid Biogenesis to Access a Skeletally Diverse Chemical Library

Song Liu, John S. Scotti, and Sergey A. Kozmin*

Department of Chemistry, University of Chicago, Chicago, Illinois 60607, United States

Supporting Information

ABSTRACT: We have developed a synthetic strategy that mimics the diversity-generating power of monoterpenoid indole alkaloid biosynthesis. Our general approach goes beyond diversification of a single natural product-like substructure and enables production of a highly diverse collection of small molecules. The reaction sequence begins with rapid and highly modular assembly of the tetracyclic indoloquinolizidine core, which can be chemoselectively processed into several additional skeletally diverse structural frameworks. The general utility of this approach was demonstrated by parallel synthesis of two representative.



demonstrated by parallel synthesis of two representative chemical libraries containing 847 compounds with favorable physicochemical properties to enable its subsequent broad pharmacological evaluation.

INTRODUCTION

Secondary metabolism creates a diverse ensemble of natural products. The power of natural product biogenesis has stimulated the development of the field of combinatorial biosynthesis, which aims to generate new collections of molecules that resemble natural products.¹ However, because of the difficulty of re-engineering natural product biosynthesis pathways, such strategies have yielded only a limited number of new compounds. While a number of synthetic strategies to produce natural product-like libraries have been developed,² including biomimetic approaches,³ they typically represent diversification of a specific natural product structure and provide limited skeletal diversity. Here we describe our design and execution of a synthetic strategy that reaches beyond diversification of a single natural product-like substructure and enables production of a collection of skeletally diverse small molecules for subsequent pharmacological probe discovery.

The divergent nature of monoterpenoid indole alkaloid biosynthesis provides a wide range of natural products based on two relatively simple precursors, including tryptamine 1a and secologanin 2 (Figure 1A).⁴ The initial condensation between 1a and 2 entails a Pictet-Spengler cyclization to give strictosidine 3.5 Subsequent processing of this biosynthetic intermediate enables branching into several additional pathways. Deglycosidation, followed by intramolecular reductive amination and indole oxidation yields hydroxy indolenine 4, which can rearrange into spirocyclic geissoschizine oxindole 5.6 Alternatively, strictosidine 3 can undergo lactamization, followed by putative oxidative cleavage of the indole moiety to give ketoamide 6. Subsequent transannular cyclization produces pumiloside 7, which is ultimately converted into camptothecin.⁷ Such representative examples have demonstrated how changes in oxidation states at specific positions of a

polycyclic molecular framework could lead to a highly controlled fragmentation and progressive diversification of a complex skeletal architecture. While recent studies demonstrate that such biosynthetic pathways could be attractive for the production of unnatural indole alkaloids, only a small number of new compounds have been prepared using this approach, either by feeding unnatural tryptamine derivatives or reengineering the substrate specificity of biosynthetic enzymes.^{8,9}

Our synthetic strategy to a skeletally diverse chemical library, which is reminiscent of monoterpenoid indole alkaloids, is depicted in Figure 1B. First, we aimed to develop an efficient and highly diastereoselective entry into a series of substituted indoloquinolizidines 10, representing the skeletal branching point. Those compounds can be readily assembled starting with appropriate tryptamines 1, 1,3-dicarbonyls 8 and itaconic anhydride 9. The next challenge was to identify reaction conditions that would efficiently reshape polycyclic molecular architecture of 10 in the absence of biosynthetic enzymes. We considered a series of possible transformations of this key synthetic intermediate into other polycyclic skeletal frameworks via chemoselective oxidation of the indole moiety, which would induce subsequent controlled fragmentation of this tetracyclic core structure. Chemoselective oxidation at the C(7) could furnish hydroxyindoline 11. Subsequent rearrangement of 11 would produce spirocyclic dihydroindolone 12. Alernatively, oxidative cleavage of indologuinolizidine 10 at the C(2)-C(7)bond would produce ring-opened ketoamide 13, which could be further elaborated into quinolone 14 via transannular aldol reaction, followed by dehydration. This approach would deliver a chemical library containing multiple core structures, which

Received: June 24, 2013 **Published:** August 12, 2013



Figure 1. (A) Biosynthetic interconversions of representative monoterpenoid indole alkaloids via intermediacy of strictosidine 3. (B) Synthetic strategy to a skeletally diverse chemical library via rapid assembly and subsequent transformations of indoloquinolizidine 10, which served as a biomimetic branching point.

could be readily decorated at several positions providing rapid access to both skeletal and peripheral diversity.

RESULTS AND DISCUSSION

We began our studies by examining construction of the indoloquinolizidine skeleton **10**. In order to rapidly assemble the tetracyclic molecular framework, we took advantage of the known propensity of vinylogous carbamates to undergo *N*-acyliminium ion cyclizations when subjected to either unsaturated acid chlorides or acid anhydrides.¹⁰ Condensation of tryptamine **1b** with ketoamide **8a** gave the corresponding vinylogous urea **15a**, which was next treated with itaconic anhydride **9** to give indoloquinolizidine **10a** in 90% yield (Scheme 1). The relative stereochemistry of **10a** was established by X-ray crystallography. This process entailed a 1,4-addition of the vinylogous urea derived from **1b** and **8a** to a Michael acceptor **9**, followed by anhydride opening and *N*-acyliminium ion cyclization. Following solvent and temperature optimization studies, we found that this transformation





proceeded with excellent diastereoselectivity when the reaction was conducted in 1,4-dioxane at 40 $^\circ \rm C.$

We next examined the generality of this transformation and found that a range of substituted tryptamines and 1,3ketoamides can be successfully employed to produce the corresponding indoloquinolizidines in high yields and excellent diastereoselectivity. A representative example of these studies is shown in Scheme 2. Treatment of 5-chloro-tryptamine **1b** with





ketoamide **8b** cleanly produced vinylogous urea **15b**, which was efficiently cyclized to afford indoloquinolizidine **10b** in a one-flask operation upon treatment with **9** and HCl in dioxane at 40 °C. The structure of **10b** was also confirmed by X-ray crystallography.

Having established a highly efficient and general indoloquinolizidine synthesis, we next examined its application to a larger set of compounds based on this tetracyclic platform (Figure 2). We prepared a 480-member library 17 starting with five tryptamines 1 and eight ketoamides 8 to deliver 40 indoloquinolizidine-containing carboxylic acids 10. Each of



Figure 2. Synthesis of 476-member library of indoloquinolizidines. The assembly process entailed two chromatographic steps, including initial purification of each of the 40 acids 10 and subsequent LC–MS purification of all final library members 17.

the initially produced acids 10 would be next diversified with 12 amines 16 to deliver the final library 17. Selection of each of the building blocks shown in Figure 2 was guided by Accelrys Pipeline Pilot, which enabled virtual enumeration of the target library and estimation of various molecular properties in silico. Specifically, during the design process, we applied appropriate filters to select building blocks that would produce the target library with molecular weight below 600 and calculated logarithmic value of *n*-octanol/water partition coefficient (c log P) within a range of 1-5. The synthesis began with condensations of five tryptamines 1 with eight ketoamides 8, followed by reactions of the resulting vinylogous ureas 15 with anhydride 9 to afford 40 indologuinolizidine-containing carboxylic acids 10. Each of the acids 10 was obtained as a single diastereomer approximately on a 500 mg scale in a single-flask operation starting with the corresponding tryptamines 1 and ketoamides 8. To ensure their high chemical purity, all pyrrolidinone-containing carboxylic acids 10 were purified by conventional chromatography. The second stage of the synthesis entailed coupling of each of the acids 10 with 12 amines 16. Following examination of several amide-coupling protocols, we established that this transformation can be efficiently accomplished using O-(7-azabenzotriazol-1-yl)-N, N, N', N'-tetramethyluronium hexafluorophosphate (HATU) and N,N-diisopropylethylamine (DIPEA) in DMF. Preparative LC-MS purification of each final library member 17 established that 476 compounds were successfully produced on 20 mg scale in 65% average yield and >90% chemical purity.

Next, we examined a range of conditions that could promote chemoselective oxidation at the C(7) position of the indole moiety. This process emulates a putative conversion of strictosidine to hydroxy indolenine 4 (Figure 1A). We found that the use of oxaziridine 18^{11} efficiently promoted this conversion. A representative alcohol 11a was obtained as a single diastereomer by treatment of the corresponding indoloquinolizidine 17a with oxaziridine 18 at ambient temperature in dichloromethane (Scheme 3). The relative stereochemistry of 11a was determined by X-ray crystallog-raphy.

Scheme 3. Synthesis of Hydroxyindoline 11a



We found that treatment of hydroxy indolenine **11b** with a dilute solution of a strong Brønsted acid triggered efficient conversion to a single reaction product. X-ray crystallography unambiguously established the structure of spirocyclic dihydroindolone **12a**,¹² which was produced with complete diastereoelectivity (Scheme 4). While this transformation was analogous to the putative biosynthetic conversion of hydroxy indolenine **4** into geissoschizine oxindole **5** (Figure 1A), the course of the rearrangement was altered, presumably because of the higher propensity of **11b** for protonation at the imine moiety, which induced subsequent [1,2]-shift with a concomitant cleavage of the C(6)–C(7) bond.

We next turned our attention to oxidative cleavage of indoloquinolizidine at the C(2)-C(7) bond. Biosynthetically, a similar transformation has been proposed to enable conversion of strictosidine 3 into pumiloside 7 via the intermediacy of 6 (Figure 1A). Among several oxidants examined, *m*-CPBA was found to be the most efficient reagent in promoting this oxidative cleavage. In a representative case, treatment of indoloquinolizidine 17b with *m*-CPBA at ambient temperature furnished the desired ring-fragmented tricylic ketoamide 13a in 71% yield (Scheme 5).

We also found that subjection of ketoamide **13b** to KOH in EtOH triggered efficient transannular aldol condensation with a concomitant dehydration.¹³ This transformation produced tetracylic quinolone **14a**, which was isolated in 53% yield

Article

Scheme 4. Synthesis of Spirocyclic Dihydroindolone 12a



Scheme 5. Synthesis of Tricyclic Ketone 13a



Scheme 6. Synthesis of Pyrroloquinolone 14a



(Scheme 6). Combination of NMR spectroscopy with X-ray crystallography once again provided unambiguous structural assignments of this compound.

Having established an efficient skeletal diversification strategy, we aimed to apply this biogenesis-guided approach to parallel assembly of a new chemical library (Figure 3). The main challenge was to ensure that iterative processing and parallel diversification of each of the core structures within the library would be efficient and compatible with a range of synthetic precursors required for library synthesis. The parallel assembly process began with condensations of four readily available tryptamines 1 with each of the two ketoamides 8, followed by treatment of the resulting vinylogous ureas with itaconic anhydride 9 to produce eight possible carboxylic acids 10 (75% average yield). The next stage of the assembly process entailed HATU-promoted coupling of 8 carboxylic acids 10 with 12 amines 16, which successfully produced 96 possible amides 17 (74% average yield after parallel LC-MS purification). The first skeletal transformation entailed oxidation of indoloquinolizidines 17 with oxaziridine 18 to give the corresponding hydroxy indolenines, which displayed high propensity to undergo subsequent acid-catalyzed rearrangement and were used directly to produce the corresponding spirocyclic dihydroindolones 12. We determined that 83 such reactions proceeded successfully to give final products 12 as single detectable diastereomers in high chemical purity (>90%) following LC–MS purification. The next skeletal transformation was accomplished by treatment of 96 indologuinolizidines 17 with m-CPBA, which successfully produced 96 required ketoamides 13 (>90% purity by LC-MS). The final skeletal elaboration entailed base-promoted transannular aldol cyclization, followed by dehydration, which



Figure 3. Synthesis of 371-member skeletally diverse small-molecule library. The assembly process mimics skeletal branching of monoterpenoid alkaloid biosynthesis and enables efficient conversion of the initially produced set of 96 indoloquinolizidines 17 into collections of skeletally distinct polycyclic products 12, 13, and 14. The figure shows structures of building blocks used for library generation, depicts reaction conditions and scales, average yields, molecular weights, chemical purities as well as log P distributions.

was applied to all ketoamides 13 to give 96 tetracyclic quinolones 14 in 64% average yield (>90% purity by LC-MS). This operation completed the construction of 371-member chemical library. All final compounds were found to exhibit excellent chemical stability and highly favorable physicochemical properties (Figure 3). This effort validated the efficiency of our approach, as well as the compatibility of this biomimetic reaction sequence with a range of building blocks required for high-throughput organic synthesis.

Newly generated chemical libraries have been subjected to broad biological evaluation conducated at the NIH Molecular Libraries Probe Production Centers Network (MLPCN). Initial screening efforts resulted in identification of several compounds with promising activity in several assays including inhibition of fatty acid synthase thioesterase domain, antagonism of Smad3specific TGF- β signaling pathway, and disruption of RIN1/ABL interaction.¹⁴ In addition, our own screening effort recently identified a new chemical inducer of mitochondrial permeability transition.¹⁵ These preliminary studies validate the general utility of this library for discovery of bioactive chemical probes and set the stage for subsequent pharmacological studies.

CONCLUSION

High-throughput synthesis of skeletally diverse small-molecule libraries remains challenging despite the notable progress achieved to date.¹⁶ As natural products continue to stimulate innovation in chemical biology and drug discovery,¹⁷ interest is growing in developing efficient approaches to chemical libraries that would approach the level of skeletal diversity found in nature. Inspired by the diversity-generating power of

monoterpenoid alkaloid biosynthesis, we present an efficient synthetic strategy to a chemical library that mimics the skeletal diversity found in nature. After initial assembly of 476-member library of indoloquinolizidines, the core structure was transformed into three additional frameworks, generating a new library of 371 compounds that feature both skeletal and peripheral diversity. Preliminary high-throughout screening of this compound collection resulted in the identification of several useful chemical probes, setting the stage for their subsequent detailed pharmacological evaluation.

EXPERIMENTAL SECTION

General Experimental Methods. Common HPLC and ACS grade organic solvents were purchased and used without further purification. Dichloromethane and tetrahydrofuran were purified by distillation. Commercially available reagents were used without further purification. Reactions were monitored by thin layer chromatography (TLC) using precoated silica gel plates. ¹H NMR and ¹³C NMR spectra were recorded on ¹H 400, 500 MHz and ¹³C 100, 125 MHz spectrometers using residual solvent peaks as an internal standard. High-resolution mass spectra were recorded with Q-TOF Ultima tandem quadrupole/Time-of-Flight instrument. LC-MS purification and analyses were performed using a LC-MS system consisting of a binary gradient module, a HPLC pump, a quadrupole mass spectrometer, a fluidics organizer, a sample manager, a dual channel UV-vis detector, and an evaporative light scattering detector. Fractions were collected by ES+ MS detection of product ion. Purity was determined by area under curve of either UV(214 nm) or ELS trace

Indoloquinolizidine 10a. A solution of 5-chlorotryptamine **1b** (204.4 mg, 1.05 mmol) in $CHCl_3$ (3 mL) was treated with ketoamide **8a** (176.0 mg, 1.04 mmol) and 4 Å molecular sieves (0.25 g). The

reaction mixture was heated at 50 °C overnight and filtered, and the filtrate was concentrated in vacuo. A solution of crude enamide in 1,4dioxane (3 mL) was treated with itaconic anhydride 9 (122.2 mg, 1.04 mmol). The reaction mixture was heated at 40 °C overnight, followed by dropwise addition of HCl (4 M in 1,4-dioxane, 0.39 mL, 1.56 mmol). Stirring was continued for 3 h at the same temperature before the solvent was removed in vacuo. The crude product was purified by flash chromatography on silica gel (elution with CHCl2:CHCl3:MeOH = 2:2:1 + 2% formic acid) to give indoloquinolizidine 10a (428.6 mg, 90%). White amorphous solid: ¹H NMR (400 MHz, CDCl₃) δ 0.70– 0.75 (m, 1H), 1.10-1.12 (m, 1H), 1.39-1.45 (m, 3H), 1.48-1.51 (m, 1H), 1.78 (d, 1H, J = 14.4 Hz), 1.91 (s, 3H), 2.58 (dt, 1H, J = 12.8, 12.0 Hz), 2.65-2.68 (m, 2H), 2.78-2.91 (m, 2H), 2.97-3.07 (m, 4H), 3.31-3.41 (m, 2H), 3.58-3.66 (m, 1H), 5.04 (dd, 1H, J = 12.4, 1.6 Hz), 7.03 (d, 1H, J = 8.8 Hz), 7.24 (d, 1H, J = 8.0 Hz), 7.43 (s, 1H), 9.08 (s, 1H), 11.3 (br s, 1H); 13 C NMR (125 MHz, CDCl₃) δ 21.0, 23.9, 25.3, 25.9, 27.3, 34.6, 37.5, 37.7, 43.6, 45.9, 47.2, 59.7, 108.7, 112.0, 117.7, 122.3, 125.0, 126.9, 134.2, 137.3, 170.6, 171.7, 175.8; HRMS (ESI) calculated for $C_{24}H_{29}N_3O_4Cl$ 458.1847 [M + H]⁺, found 458.1838. Single crystals for X-ray analysis were obtained by slow evaporation of saturated solution method. CCDC number is 791451.

Indologuinolizidine 10b. A solution of 5-chlorotryptamine 1b (205.5 mg, 1.06 mmol) in CHCl₃ (4 mL) was treated with ketoamide 8b (273.1 mg, 1.05 mmol) and 4 Å molecular sieves (0.25 g). The reaction mixture was heated at 50 °C overnight and filtered, and the filtrate was concentrated in vacuo. A solution of crude enamide in 1,4dioxane (4 mL) was treated with itaconic anhydride 9 (114.7 mg, 1.02 mmol). The reaction mixture was heated at 40 °C overnight, followed by dropwise addition of HCl (4 M in 1,4-dioxane, 0.40 mL, 1.60 mmol). Stirring was continued for 3 h at the same temperature before the solvent was removed in vacuo. The crude product was purified by flash chromatography on silica gel (elution with CHCl₂:CHCl₃:MeOH = 2:2:1 + 2% formic acid) to give indoloquinolizidine 10b (322.9 mg, 56%). White amorphous solid: ¹H NMR (500 MHz, CDCl₃) δ 0.28 (br s, 1H), 0.92 (br s, 1H), 1.16 (s, 1H), 1.37 (s, 1H), 1.68 (dd, 1H, J = 7.5, 5.5 Hz), 2.30 (t, 1H, J = 11.3 Hz), 2.41 (td, 1H, J = 14.0, 4.4 Hz), 2.52 (td, 1H, J = 12.6, 4.1 Hz), 2.61-2.89 (m, 7H), 2.93-3.18 (m, 6H), 3.40 (dd, 1H, J = 11.2, 7.9 Hz), 3.51–3.60 (m, 1H), 5.26 (d, 1H, J = 8.6), 7.06 (dd, 3H, J = 11.5, 4.5 Hz), 7.10 (t, 1H, J = 7.3 Hz), 7.18 (t, 2H, J = 7.5 Hz), 7.30 (d, 1H, J = 8.6 Hz), 7.45 (s, 1H), 9.67 (s, 1H), 11.45 (br s, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 20.9, 23.9, 25.4, 25.9, 27.5, 31.3, 34.6, 37.0, 37.5, 40.9, 43.9, 47.4, 48.2, 62.7, 110.8, 112.4, 117.9, 122.7, 125.3, 126.1, 127.1, 128.3, 128.5, 134.7, 134.9, 141.2, 171.3, 171.6, 176.8; HRMS (ESI) calculated for $C_{31}H_{35}N_{3}O_{4}Cl$ 548.2316 [M + H]⁺, found 548.2319. Single crystals for X-ray analysis were obtained by slow evaporation of saturated solution method. CCDC number is 900900.

Indoloquinolizidine 10c. A solution of tryptamine 1a (170.9 mg, 1.05 mmol) in CHCl₃ (3 mL) was treated with 1-morpholinobutane-1,3-dione 8e (181.3 mg, 1.04 mmol) and 4 Å molecular sieves (0.25 g). The reaction mixture was heated at 50 °C overnight and filtered, and the filtrate was concentrated in vacuo. To a solution of crude enamide in 1,4-dioxane (3 mL) was added itaconic anhydride 9 (122.2 mg, 1.04 mmol). The reaction mixture was heated at 40 °C overnight, followed by dropwise addition of HCl (4 M in 1,4-dioxane, 0.39 mL, 1.56 mmol). Stirring was continued for 3 h at the same temperature before the solvent was removed in vacuo. The crude product was purified by flash chromatography on silica gel (elution with $EtOAc:CHCl_3:MeOH = 5:5:1 + 2\%$ formic acid) to give indoloquinolizidine 10c (429.2 mg, 97%). White amorphous solid: ¹H NMR (500 MHz, CDCl₃) δ 1.77 (dt, 1H, J = 13.5, 3.5 Hz), 1.90 (s, 3H), 2.61 (dt, 1H, J = 13.5, 11.5 Hz), 2.73–2.82 (m, 4H), 2.87–2.94 (m, 2H), 2.99-3.03 (m, 1H), 3.04-3.09 (m, 2H), 3.12-3.16 (m, 1H), 3.37 (dd, 1H, J = 13.0, 5.0 Hz), 3.49 (d, 2H, J = 8.0 Hz), 3.58 (t, 1H, J = 7.5 Hz), 3.61–3.66 (m, 1H), 5.05 (d, 1H, J = 11.5 Hz), 7.03–7.07 (m, 2H), 7.27 (d, 1H, J = 7.5 Hz), 7.47 (d, 1H, J = 8.0 Hz), 8.87 (s, 1H), 10.65 (br s, 1H); 13 C NMR (125 MHz, CDCl₃) δ 21.0, 21.2, 27.3, 34.5, 37.6, 37.7, 42.5, 45.9, 46.2, 59.6, 65.8, 66.3, 109.3, 111.0, 118.3, 119.6, 122.3, 125.9, 135.5, 135.8, 170.6, 172.3, 175.6; HRMS (ESI) calculated for $C_{23}H_{28}N_3O_5$ 426.2029 $[M+H]^+$, found 426.2016.

Indoloquinolizidine 10d. It was prepared from tryptamine 1a, ketoamide 8a, and itaconic anhydride 9 in 339.6 mg, 80% yield by following the same protocol of **10c**. White amorphous solid: ¹H NMR (500 MHz, CDCl₃) δ 0.82–0.85 (m, 1H), 1.16–1.18 (m, 1H), 1.34–1.38 (m, 1H), 1.43–1.45 (m, 2H), 1.47–1.51 (m, 1H), 1.78 (dt, 1H, *J* = 14.0, 4.0 Hz), 1.94 (s, 3H), 2.59 (dt, 1H, *J* = 13.5, 11.0 Hz), 2.72–2.76 (m, 1H), 2.77–2.78 (m, 1H), 2.83 (dd, 1H, *J* = 17.0, 5.0 Hz), 2.89–2.97 (m, 2H), 3.01–3.06 (m, 1H), 3.07–3.18 (m, 2H), 3.31 (dd, 1H, *J* = 13.5, 5.0 Hz), 3.43–3.46 (m, 1H), 3.59–3.63 (m, 1H), 5.05 (dd, 1H, *J* = 12.5, 2.5 Hz), 7.07 (t, 1H, *J* = 7.5 Hz), 7.13 (t, 1H, *J* = 7.5 Hz), 7.29 (d, 1H, *J* = 8.5 Hz), 7.47 (d, 1H, *J* = 8.0 Hz), 8.85 (s, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 21.2, 21.4, 24.1, 25.5, 26.2, 27.5, 34.9, 37.8, 38.2, 43.5, 46.0, 47.3, 60.0, 109.0, 111.1, 118.3, 119.5, 122.2, 126.0, 135.9, 136.0, 171.0, 171.8, 175.6; HRMS (ESI) calculated for C₂₄H₃₀N₃O₄ 424.2236 [M + H]⁺, found 424.2223.

Indoloquinolizidine 10e. It was prepared from tryptamine 1a, ketoamide 8g, and itaconic anhydride 9 in 320.4 mg, 63% yield by following the same protocol of 10c. Pale yellow amorphous solid: ¹H NMR (500 MHz, CDCl₃) δ 0.84–0.88 (m, 1H), 0.90–0.99 (m, 2H), 1.31–1.36 (m, 1H), 1.42–1.50 (m, 4H), 1.62–1.67 (m, 1H), 1.68–1.74 (m, 3H), 2.14–2.20 (m, 1H), 2.21–2.26 (m, 1H), 2.58–2.60 (m, 1H), 2.70–2.74 (m, 4H), 2.75–2.80 (m, 1H), 2.82–2.86 (m, 1H), 2.89–2.95 (m, 2H), 3.01–3.06 (m, 1H), 3.10 (dd, 1H, *J* = 17.0, 5.0 Hz), 3.29–3.35 (m, 3H), 3.48–3.51 (m, 1H), 3.60–3.63 (m, 1H), 5.22 (dd, 1H, *J* = 12.0, 2.5 Hz), 7.04–7.10 (m, 2H), 7.28 (d, 1H, *J* = 7.0 Hz), 7.50 (d, 1H, *J* = 7.0 Hz), 8.95 (s, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 21.0, 25.1, 25.1, 27.4, 31.1, 32.5, 32.8, 33.6, 34.2, 37.1, 40.3, 40.7, 42.5, 46.1, 47.9, 62.5, 65.5, 66.1, 110.9, 118.3, 119.7, 122.3, 126.0, 133.6, 136.0, 171.2, 171.9, 176.0; HRMS (ESI) calculated for C₂₉H₃₈N₃O₅ 508.2811 [M + H]⁺, found 508.2798.

Indoloquinolizidine 10f. It was prepared from tryptamine 1a, ketoamide 8i, and itaconic anhydride 9 in 211.5 mg, 55% yield by following the same protocol of 10c. White amorphous solid: ¹H NMR (500 MHz, CDCl₃) δ 1.81 (dt, 1H, *J* = 13.0, 3.5 Hz), 1.90 (s, 3H), 2.58 (s, 3H), 2.72–2.86 (m, 3H), 2.87–2.93 (m, 4H), 2.97–3.05 (m, 3H), 3.36 (dd, 1H, *J* = 13.0, 4.5 Hz), 5.06 (d, 1H, *J* = 11.0 Hz), 7.06 (t, 1H, *J* = 7.5 Hz), 7.11 (t, 1H, *J* = 7.5 Hz), 7.32 (d, 1H, *J* = 8.0 Hz), 7.47 (t, 1H, *J* = 7.5 Hz), 9.01 (s, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 20.9, 21.0, 26.6, 34.6, 36.1, 37.2, 37.6, 37.7, 46.0, 59.5, 109.0, 111.1, 118.0, 119.2, 121.9, 125.7, 135.6, 135.9, 170.6, 173.5, 175.3; HRMS (ESI) calculated for C₂₁H₂₆N₃O₄ 384.1923 [M + H]⁺, found 384.1923.

Indoloquinolizidine 10g. It was prepared from tryptamine **1b**, ketoamide **8i**, and itaconic anhydride **9** in 263.7 mg, 63% yield by following the same protocol of **10c**. Pale yellow amorphous solid: ¹H NMR (500 MHz, CDCl₃) δ 1.79 (dt, 1H, *J* = 13.0, 3.5 Hz), 1.88 (s, 3H), 2.52–2.57 (m, 1H), 2.60 (s, 3H), 2.64–2.79 (m, 3H), 2.80–2.85 (m, 1H), 2.89 (s, 3H), 2.97–3.01 (m, 2H), 3.32 (dd, 1H, *J* = 13.0, 5.0 Hz), 5.03 (d, 1H, *J* = 11.0 Hz), 7.02 (d, 1H, *J* = 9.0 Hz), 7.21 (d, 1H, *J* = 8.5 Hz), 7.42 (s, 1H), 9.07 (s, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 21.0, 26.9, 34.7, 36.3, 37.4, 37.6, 37.9, 46.3, 59.6, 108.9, 112.2, 117.8, 122.3, 125.2, 126.9, 134.3, 137.3, 170.7, 173.6, 175.9; HRMS (ESI) calculated for C₂₁H₂₅N₃O₄Cl 418.1534 [M + H]⁺, found 418.1534.

Indoloquinolizidine 10h. It was prepared from tryptamine 1b, ketoamide **8e**, and itaconic anhydride **9** in 299.3 mg, 65% yield by following the same protocol of **10c**. Pale yellow amorphous solid: ¹H NMR (500 MHz, CDCl₃) δ 1.76 (dt, 1H, *J* = 13.5, 3.5 Hz), 1.91 (s, 3H), 2.58 (dt, 1H, *J* = 13.5, 11.0 Hz), 2.70–2.75 (m, 3H), 2.85–2.90 (m, 2H), 2.98–3.07 (m, 3H), 3.17–3.22 (m, 2H), 3.33 (dd, 1H, *J* = 13.0, 5.0 Hz), 3.47–3.53 (m, 2H), 3.58–3.64 (m, 2H), 5.04 (d, 1H, *J* = 11.5 Hz), 7.02 (dd, 1H, *J* = 8.5, 2.0 Hz), 7.18 (d, 1H, *J* = 9.0 Hz), 7.44 (d, 1H, *J* = 1.5 Hz), 8.92 (s, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 21.1, 27.4, 34.6, 37.7, 42.6, 46.0, 46.3, 59.7, 66.1, 66.4, 109.2, 112.0, 118.0, 122.7, 125.4, 127.0, 134.2, 137.1, 170.6, 172.3, 175.8; HRMS (ESI) calculated for C₂₃H₂₇N₃O₅Cl 460.1639 [M + H]⁺, found 460.1639.

Indoloquinolizidine 10i. It was prepared from tryptamine **1***c*, ketoamide **8***f*, and itaconic anhydride **9** in 250.6 mg, 49% yield by following the same protocol of **10***c*. Pale yellow amorphous solid: ¹H

NMR (500 MHz, CDCl₃) δ 0.64 (t, 3H, *J* = 7.0 Hz), 1.11 (t, 3H, *J* = 7.0 Hz), 1.75–1.77 (m, 1H), 1.90 (s, 3H), 2.57–2.65 (m, 3H), 2.69–2.75 (m, 2H), 2.77–2.86 (m, 3H), 2.89–2.96 (m, 8H), 3.01–3.07 (m, 4H), 3.22 (dd, 1H, *J* = 13.5, 4.5 Hz), 3.26 (dd, 1H, *J* = 13.5, 7.0 Hz), 3.38–3.43 (m, 1H), 5.03 (d, 1H, *J* = 12.5 Hz), 6.95 (d, 1H, *J* = 8.0 Hz), 7.09 (s, 1H), 7.37 (d, 1H, *J* = 8.0 Hz), 8.42 (s, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 12.9, 14.2, 21.1, 21.2, 27.6, 31.7, 34.9, 35.5, 36.0, 37.2, 37.5, 38.2, 41.2, 42.6, 46.5, 59.5, 108.8, 110.5, 118.3, 120.3, 124.4, 135.6, 135.7, 136.1, 170.8, 172.6, 173.0, 175.5; HRMS (ESI) calculated for C₂₈H₃₉N₄O₅ 511.2920 [M + H]⁺, found 511.2913.

Indoloquinolizidine 10j. It was prepared from tryptamine 1e, ketoamide 8a, and itaconic anhydride 9 in 390.9 mg, 86% yield by following the same protocol of 10c. Pale yellow amorphous solid: ¹H NMR (500 MHz, CDCl₃) δ 0.88–0.92 (m, 1H), 1.19–1.24 (m, 1H), 1.43–1.50 (m, 4H), 1.76 (dt, 1H, *J* = 13.0, 4.0 Hz), 1.95 (s, 3H), 2.55–2.62 (m, 1H), 2.76–2.78 (m, 2H), 2.82–2.95 (m, 3H), 3.02–3.05 (m, 1H), 3.14–3.17 (m, 2H), 3.27 (dd, 1H, *J* = 13.5, 5.0 Hz), 3.42–3.49 (m, 1H), 3.63–3.66 (m, 1H), 3.83 (s, 3H), 5.05 (dt, 1H, *J* = 12.0, 2.0 Hz), 6.80 (dd, 1H, *J* = 9.0, 2.5 Hz), 6.92 (d, 1H, *J* = 2.5 Hz), 7.18 (d, 1H, *J* = 8.5 Hz), 8.64 (s, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 21.2, 21.5, 24.2, 25.6, 26.3, 27.7, 35.0, 37.8, 38.5, 43.5, 46.0, 47.3, 55.9, 60.1, 100.3, 108.7, 111.8, 112.3, 126.3, 130.9, 136.7, 154.1, 171.1, 171.7, 175.0; HRMS (ESI) calculated for C₂₅H₃₂N₃O₅ 454.2342 [M + H]⁺, found 454.2334.

Indoloquinolizidine 10k. It was prepared from tryptamine 1e, ketoamide 8b, and itaconic anhydride 9 in 381.2 mg, 70% yield by following the same protocol of 10c. Pale yellow amorphous solid: ¹H NMR (500 MHz, CDCl₃) δ 0.39–0.42 (m, 1H), 0.92–0.94 (m, 1H), 1.21–1.28 (m, 3H), 1.39–1.42 (m, 1H), 1.73–1.75 (m, 1H), 2.37–2.43 (m, 2H), 2.53–2.58 (m, 1H), 2.62–2.65 (m, 1H), 2.76–2.80 (m, 2H), 2.88–2.92 (m, 3H), 3.03–3.07 (m, 3H), 3.18–3.23 (m, 2H), 3.37 (dd, 1H, *J* = 12.5, 6.0 Hz), 3.62–3.64 (m, 1H), 3.84 (s, 3H), 5.29 (d, 1H, *J* = 8.5 Hz), 6.80 (d, 1H, *J* = 9.0 Hz), 6.96 (s, 1H), 7.07–7.14 (m, 3H), 7.19–7.26 (m, 3H), 8.94 (s, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 21.0, 23.9, 25.3, 25.7, 27.5, 31.3, 34.6, 37.0, 37.4, 40.9, 43.6, 47.2, 48.1, 55.9, 62.6, 100.3, 110.7, 111.9, 112.3, 126.0, 126.4, 128.2, 128.4, 131.3, 134.1, 141.3, 154.0, 171.4, 171.5, 176.2; HRMS (ESI) calculated for C₃₂H₃₈N₃O₅ 544.2811 [M + H]⁺, found 544.2794.

Indoloquinolizidine 10I. It was prepared from tryptamine **1e**, ketoamide **8i**, and itaconic anhydride **9** in 285.8 mg, 69% yield by following the same protocol of **10c**. Pale yellow amorphous solid: ¹H NMR (500 MHz, CDCl₃) δ 1.79 (dt, 1H, *J* = 13.0, 3.5 Hz), 1.93 (s, 3H), 2.52–2.59 (m, 1H), 2.72 (s, 3H), 2.74–2.76 (m, 2H), 2.82–2.85 (m, 1H), 2.86–2.91 (m, 2H), 2.92 (s, 3H), 3.01–3.06 (m, 1H), 3.27 (dd, 1H, *J* = 13.5, 5.0 Hz), 3.83 (s, 3H), 5.06 (d, 1H, *J* = 11.0 Hz), 6.81 (dd, 1H, *J* = 8.5, 2.5 Hz), 6.92 (d, 1H, *J* = 2.5 Hz), 7.19 (d, 1H, *J* = 8.5 Hz), 8.62 (s, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 21.2, 26.9, 34.9, 36.2, 37.5, 37.7, 38.2, 46.2, 55.9, 59.8, 100.3, 108.8, 112.0, 112.1, 126.2, 131.0, 136.6, 154.0, 170.9, 173.6, 175.8; HRMS (ESI) calculated for C₂₂H₂₈N₃O₅ 414.2029 [M + H]⁺, found 414.2037.

Indoloquinolizidine 10m. It was prepared from tryptamine 1e, ketoamide 8e, and itaconic anhydride 9 in 429.1 mg, 94% yield by following the same protocol of 10c. Pale yellow amorphous solid: ¹H NMR (500 MHz, CDCl₃) δ 1.75 (dt, 1H, *J* = 13.0, 4.0 Hz), 1.89 (s, 3H), 2.58 (dt, 1H, *J* = 13.5, 11.0 Hz), 2.71–2.82 (m, 4H), 2.87–2.91 (m, 1H), 2.98–3.07 (m, 3H), 3.12–3.19 (m, 2H), 3.35 (dd, 1H, *J* = 13.0, 5.0 Hz), 3.48–3.52 (m, 2H), 3.58–3.65 (m, 2H), 3.76 (s, 3H), 5.04 (d, 1H, *J* = 12.0 Hz), 6.71 (dd, 1H, *J* = 8.5, 2.0 Hz), 6.92 (d, 1H, *J* = 2.0 Hz), 7.15 (d, 1H, *J* = 8.5 Hz), 8.69 (s, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 21.1, 21.3, 27.4, 34.6, 37.7, 37.8, 42.5, 46.0, 46.3, 55.8, 59.7, 66.0, 66.4, 100.6, 109.1, 111.7, 112.2, 126.3, 131.0, 136.4, 154.1, 170.7, 172.3, 175.7; HRMS (ESI) calculated for C₂₄H₃₀N₃O₆ 456.2135 [M + H]⁺, found 456.2134.

Indoloquinolizidine 10n. It was prepared from tryptamine 1f, ketoamide 8i, and itaconic anhydride 9 in 310.6 mg, 78% yield by following the same protocol of **10c**. Pale yellow amorphous solid: ¹H NMR (500 MHz, CDCl₃) δ 1.79 (dt, 1H, *J* = 13.0, 3.5 Hz), 1.99 (s, 3H), 2.43 (s, 3H), 2.59 (dt, 1H, *J* = 13.5, 10.5 Hz), 2.77 (s, 3H), 2.79 – 2.81 (m, 2H), 2.86–2.89 (m, 2H), 2.95 (s, 3H), 2.96–3.00 (m, 1H), 3.06–3.09 (m, 1H), 3.24 (dd, 1H, *J* = 13.5, 5.0 Hz), 5.05 (dd, 1H, *J* =

13.0, 3.0 Hz), 6.98 (d, 1H, *J* = 7.5 Hz), 7.04 (t, 1H, *J* = 7.5 Hz), 7.35 (d, 1H, *J* = 7.5 Hz), 8.55 (s, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 16.5, 21.1, 26.6, 34.8, 35.5, 36.0, 37.3, 37.6, 37.9, 46.0, 59.7, 109.6, 115.9, 119.6, 120.3, 122.6, 125.4, 135.2, 135.6, 170.7, 173.6, 175.5; HRMS (ESI) calculated for C₂₂H₂₈N₃O₄ 398.2080 [M + H]⁺, found 398.2081.

Indoloquinolizidine 10o. It was prepared from tryptamine 1**f**, ketoamide **8e**, and itaconic anhydride **9** in 339.2 mg, 77% yield by following the same protocol of **10c**. Pale yellow amorphous solid: ¹H NMR (500 MHz, CDCl₃) δ 1.76 (dt, 1H, *J* = 13.0, 4.0 Hz), 1.98 (s, 3H), 2.42 (s, 3H), 2.60 (dt, 1H, *J* = 13.5, 10.5 Hz), 2.75–2.80 (m, 3H), 2.93–3.03 (m, 3H), 3.09–3.14 (m, 1H), 3.25–3.31 (m, 2H), 3.33–3.38 (m, 1H), 3.39–3.43 (m, 1H), 3.49–3.52 (m, 1H), 3.58–3.61 (m, 2H), 3.75 (dt, 1H, *J* = 13.0, 5.0 Hz), 5.03 (dd, 1H, *J* = 12.0, 3.0 Hz), 6.98 (d, 1H, *J* = 7.0 Hz), 7.04 (t, 1H, *J* = 7.0 Hz), 7.34 (d, 1H, *J* = 7.5 Hz), 8.48 (s, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 16.6, 21.2, 21.4, 27.5, 34.8, 37.8, 38.1, 42.2, 46.0, 46.2, 59.8, 66.4, 66.5, 109.7, 116.0, 119.9, 120.1, 122.9, 125.4, 135.2, 135.3, 170.8, 172.4, 174.9; HRMS (ESI) calculated for C₂₄H₃₀N₃O₅ 440.2185 [M + H]⁺, found 440.2183.

Indologuinolizidine 17a. A solution of indologuinolizidine 10f (15.3 mg, 0.040 mmol) and amine 16k (3 equiv.) in 1:1 THF-CHCl₃ (1.0 mL) was treated with EDCI (13.0 mg, 0.066 mmol), HOBt (8.1 mg, 0.060 mmol) and DIPEA (25 μ L, 0.14 mmol). The reaction mixture was stirred overnight at room temperature, and then solvent was removed in vacuo. The crude product was purified by flash chromatography on silica gel (elution with EtOAc:MeOH = 20:1) to give indoloquinolizidine 17a (12.6 mg, 68%). Pale yellow liquid: ¹H NMR (500 MHz, CD₃OD) δ 1.68 (dt, 1H, J = 14.0, 4.0 Hz), 1.85 (s, 3H), 2.47–2.54 (m, 1H), 2.55 (s, 3H), 2.69 (d, 2H, J = 6.0 Hz), 2.75– 2.77 (m, 2H), 2.95 (s, 3H), 2.97-3.01 (m, 1H), 3.05-3.08 (m, 1H), 3.50 (dd, 1H, J = 12.0, 4.5 Hz), 4.28 (d, 1H, J = 15.5 Hz), 4.34 (d, 1H, J = 15.5 Hz), 4.97 (dt, 1H, J = 10.0, 2.5 Hz), 6.21-6.22 (m, 1H), 6.30–6.31 (m, 1H), 7.01 (t, 1H, J = 8.0 Hz), 7.10 (t, 1H, J = 7.5 Hz), 7.32 (d, 1H, J = 8.0 Hz), 7.35–7.36 (m, 1H), 7.43 (d, 1H, J = 7.5 Hz); $^{13}\mathrm{C}$ NMR (125 MHz, CD_3OD) δ 22.0, 22.3, 27.5, 36.4, 36.7, 37.1, 37.7, 39.2, 40.1, 46.0, 61.5, 108.1, 110.4, 111.3, 112.3, 119.0, 120.3, 123.0, 127.6, 137.5, 138.0, 143.3, 153.1, 173.4, 173.7, 174.4; HRMS (ESI) calculated for $C_{26}H_{31}N_4O_4$ 463.2345 [M + H]⁺, found 463.2349.

Indoloquinolizidine 17c. It was prepared from indoloquinolizidine **10b** and amine **16j** in 17.9 mg, 71% yield by following the same protocol of **17a**. Pale yellow liquid: ¹H NMR (500 MHz, CDCl3) δ 0.78–0.88 (m, 1H), 1.34 (ddd, 3H *J* = 15.8, 11.8, 11.3 Hz), 1.44–1.54 (m, 2H), 1.71–1.82 (m, 2H), 1.89–1.97 (m, 1H), 2.40 (s, 7H), 2.44–2.67 (m, 6H), 2.78 (dd, 1H, *J* = 11.6, 4.8 Hz), 2.85–2.96 (m, 2H), 3.02–3.13 (m, 2H), 3.25 (t, 2H, *J* = 5.4 Hz), 3.32–3.47 (m, 5H), 3.63 (s, 1H), 5.30 (dd, 1H, *J* = 13.1, 3.9 Hz), 7.13–7.21 (m, 5H), 7.28 (s, 1H), 7.29 (d, 1H, *J* = 2.9 Hz), 7.32 (d, 1H, *J* = 2.3 Hz), 7.54 (d, 1H, *J* = 1.8 Hz), 8.92 (s, 1H); ¹³C NMR (125 MHz, CDCl3) δ 20.8, 24.3, 25.6, 26.0, 26.6, 27.3, 31.5, 35.4, 37.6, 38.3, 39.4, 40.3, 43.3, 45.0, 47.4, 48.2, 57.7, 62.4, 110.7, 112.2, 117.9, 122.4, 125.2, 126.0, 127.3, 128.3, 128.4, 134.4, 135.7, 141.4, 171.4, 171.6, 171.9; HRMS (ESI) calculated for $C_{36}H_{47}N_5O_3CI$ 632.3367 [M + H]⁺, found 632.3364.

Indoloquinolizidine 17d. It was prepared from indoloquinolizidine **10d** and amine **16c** in 17.2 mg, 77% yield by following the same protocol of **17a**. Pale yellow liquid: ¹H NMR (500 MHz, CDCl₃) *δ* 1.02–1.06 (m, 1H), 1.36–1.42 (m, 1H), 1.44–1.49 (m, 2H), 1.52–1.56 (m, 2H), 1.57 (s, 3H), 1.62 (s, 3H), 1.67 (s, 3H), 1.92–1.95 (m, 2H), 1.98 (s, 3H), 2.01–2.07 (m, 3H), 2.47–2.52 (m, 2H), 2.73–2.79 (m, 2H), 2.90–2.97 (m, 3H), 3.22 (dd, 1H, *J* = 13.5, 5.0 Hz), 3.29–3.33 (m, 2H), 3.51–3.55 (m, 1H), 3.61–3.65 (m, 1H), 3.73–3.82 (m, 2H), 5.02–5.04 (m, 1H), 5.06–5.11 (m, 2H), 5.91 (t, 1H, *J* = 4.5 Hz), 7.09 (t, 1H, *J* = 7.5 Hz), 7.16 (t, 1H, *J* = 7.5 Hz), 7.28 (d, 1H, *J* = 7.5 Hz), 7.48 (t, 1H, *J* = 8.0 Hz), 8.73 (s, 1H); ¹³C NMR (125 MHz, CDCl₃) *δ* 16.2, 17.6, 21.3, 21.8, 24.3, 25.7, 25.7, 26.4, 26.6, 26.7, 36.4, 37.4, 37.4, 39.5, 40.1, 43.3, 46.1, 47.3, 59.9, 108.8, 111.0, 118.3, 119.4, 119.9, 122.0, 123.8, 126.1, 131.7, 135.7, 136.6, 139.8, 170.7, 171.0, 172.2; HRMS (ESI) calculated for C₃₄H₄₇N₄O₃ 559.3648 [M + H]⁺, found 559.3633.

Indoloquinolizidine 17e. It was prepared from indoloquinolizidine 10d and amine 16f in 14.0 mg, 60% yield by following the same protocol of 17a. Pale yellow liquid: ¹H NMR (500 MHz, CDCl₃) δ 0.94-1.00 (m, 1H), 1.26-1.32 (m, 1H), 1.41-1.47 (m, 2H), 1.49-1.54 (m, 2H), 1.98 (s, 3H), 2.05 (ddd, 1H, J = 14.0, 5.0, 3.0 Hz), 2.49 (dt, 1H, J = 14.0, 11.0 Hz), 2.55-2.59 (m, 1H), 2.64-2.70 (m, 1H),2.78-2.82 (m, 1H), 2.92 (dd, 1H, J = 12.0, 3.5 Hz), 2.95-3.02 (m, 2H), 3.15-3.22 (m, 3H), 3.55-3.61 (m, 2H), 4.40-4.49 (m, 2H), 5.05 (ddd, 1H, J = 12.5, 4.5, 1.0 Hz), 6.72 (m, 1H), 7.09 (t, 1H, J = 7.0 Hz), 7.17 (dt, 1H, J = 7.5, 1.0 Hz), 7.29 (d, 1H, J = 8.0 Hz), 7.33 (d, 1H, J = 8.0 Hz), 7.42 (d, 1H, J = 7.5 Hz), 7.46-7.52 (m, 3H), 8.75 (s, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 21.2, 21.6, 24.3, 25.7, 26.5, 27.0, 36.2, 37.4, 39.9, 42.9, 43.3, 46.2, 47.1, 60.0, 108.8, 111.1, 118.3, 119.4, 122.1, 124.0 (J_{CF} = 273 Hz), 124.2 (J_{CF} = 3.9 Hz), 124.3 (J_{CF} = 3.6 Hz), 126.0, 129.0, 130.6 (J_{CF} = 1.4 Hz), 130.9 (J_{CF} = 32.3 Hz), 135.7, 136.4, 139.6, 170.8, 171.6, 172.0; HRMS (ESI) calculated for $C_{32}H_{36}N_4O_3F_3$ 581.2740 [M + H]⁺, found 581.2727.

Indoloquinolizidine 17f. It was prepared from indoloquinolizidine 10d and amine 16i in 17.6 mg, 89% yield by following the same protocol of 17a. Pale yellow liquid: ¹H NMR (500 MHz, CDCl₃) δ 1.05–1.09 (m, 1H), 1.37–1.39 (m, 1H), 1.44–1.48 (m, 2H), 1.50–1.56 (m, 2H), 1.70–1.75 (m, 2H), 1.97 (s, 3H), 2.02 (ddd, 1H, J = 14.0, 4.5, 3.0 Hz), 2.47 (dt, 1H, J = 14.0, 10.5 Hz), 2.54 (dd, 1H, J = 14.5, 4.5 Hz), 2.74–2.80 (m, 2H), 2.86 (dd, 1H, J = 14.5, 6.0 Hz), 2.93–2.96 (m, 2H), 3.24–3.34 (m, 8H), 3.42 (t, 2H, J = 6.0 Hz), 3.55–3.60 (m, 2H), 5.07 (ddd, 1H, J = 11.5, 3.0, 1.5 Hz), 6.35 (t, 1H, J = 5.0 Hz), 7.10 (t, 1H, J = 7.5 Hz), 7.16 (t, 1H, J = 7.5 Hz), 7.29 (d, 1H, J = 8.0 Hz), 7.48 (d, 1H, J = 8.0 Hz), 8.76 (s, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 21.2, 21.8, 24.3, 25.7, 26.5, 26.8, 29.2, 36.3, 37.3, 37.7, 40.0, 43.3, 46.0, 47.3, 58.7, 59.9, 71.3, 108.8, 111.0, 118.3, 119.3, 122.0, 126.1, 135.7, 136.6, 170.8, 171.2, 172.2; HRMS (ESI) calculated for C₂₈H₃₉N₄O₄ 495.2971 [M + H]⁺, found 495.2973.

Indoloquinolizidine 17g. It was prepared from indoloquinolizidine 10d and amine 16k in 12.5 mg, 62% yield by following the same protocol of 17a. Pale yellow liquid: ¹H NMR (500 MHz, CDCl₃) δ 0.99-1.04 (m, 1H), 1.33-1.37 (m, 1H), 1.42-1.47 (m, 2H), 1.50-1.56 (m, 2H), 1.98 (s, 3H), 2.08 (ddd, 1H, J = 14.5, 5.0, 3.5 Hz), 2.43-2.50 (m, 2H), 2.55-2.61 (m, 1H), 2.75 (ddd, 1H, J = 14.0, 2.0, 1.0 Hz), 2.90 (dd, 1H, J = 12.5, 3.0 Hz), 2.97-3.00 (m, 1H), 3.08 (dd, 1H, J = 14.0, 5.0 Hz), 3.17 (dd, 1H, J = 14.0, 5.0 Hz), 3.23–3.25 (m, 2H), 3.47-3.52 (m, 1H), 3.61-3.66 (m, 1H), 4.18 (dd, 1H, J = 15.5, 4.5 Hz), 4.55 (dd, 1H, J = 15.5, 6.5 Hz), 5.04 (ddd, 1H, J = 11.5, 3.5, 1.0 Hz), 6.14 (d, 1H, J = 3.0 Hz), 6.20 (d, 1H, J = 2.0 Hz), 6.49 (t, 1H, J = 5.0 Hz, 7.03 (d, 1H, J = 1.5 Hz), 7.10 (dt, 1H, J = 8.0, 1.0 Hz), 7.17 (dt, 1H, J = 8.0, 1.0 Hz), 7.29 (d, 1H, J = 8.0 Hz), 7.49 (d, 1H, J = 7.5 Hz), 8.75 (s, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 21.1, 21.7, 24.3, 25.7, 26.3, 26.7, 36.2, 36.4, 37.5, 39.8, 43.3, 46.1, 47.2, 60.0, 107.2, 108.9, 110.3, 111.1, 118.3, 119.4, 122.1, 126.1, 135.7, 136.5, 142.1, 151.3, 170.7, 171.1, 172.1; HRMS (ESI) calculated for C₂₉H₃₅N₄O₄ $503.2658 [M + H]^+$, found 503.2645.

Indoloquinolizidine 17h. It was prepared from indoloquinolizidine **10d** and amine **16g** in 15.0 mg, 71% yield by following the same protocol of **17a.** Pale yellow liquid: ¹H NMR (500 MHz, CDCl₃) δ 1.02–1.07 (m, 1H), 1.33–1.38 (m, 1H), 1.42–1.47 (m, 2H), 1.48–1.54 (m, 2H), 1.95–1.99 (m, 4H), 2.46 (dt, 1H, *J* = 14.0, 10.0 Hz), 2.59 (dd, 1H, *J* = 14.5, 4.5 Hz), 2.67–2.74 (m, 1H), 2.76–2.78 (m, 1H), 2.81 (dd, 1H, *J* = 15.0, 6.5 Hz), 2.89 (dd, 1H, *J* = 12.0, 4.0 Hz), 2.93–2.98 (m, 3H), 3.27–3.31 (m, 3H), 3.50–3.68 (m, 4H), 5.05 (ddd, 1H, *J* = 12.5, 4.0, 1.0 Hz), 6.71 (t, 1H, *J* = 5.0 Hz), 7.09 (t, 1H, *J* = 7.5 Hz), 7.13–7.17 (m, 3H), 7.28 (d, 1H, *J* = 8.0 Hz), 7.48 (d, 1H, *J* = 7.5, 1.5 Hz), 8.51 (dd, 1H, *J* = 5.5, 2.0 Hz), 8.75 (s, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 21.2, 21.8, 24.3, 25.7, 26.5, 26.8, 36.2, 37.0, 37.3, 38.7, 40.0, 43.2, 45.9, 47.2, 59.8, 108.9, 111.0, 118.3, 119.3, 121.6, 122.0, 123.3, 126.1, 135.6, 136.5, 136.6, 149.2, 159.4, 170.7, 171.1, 172.3; HRMS (ESI) calculated for C₃₁H₃₈N₅O₃ 528.2975 [M + H]⁺, found 528.2968.

Indoloquinolizidine 17i. It was prepared from indoloquinolizidine **10h** and amine **16s** in 15.7 mg, 63% yield by following the same protocol of **17a**. Pale yellow liquid: ¹H NMR (500 MHz, CD₃OD) δ 1.68 (dt, 1H, J = 13.5, 4.5 Hz), 1.88 (s, 3H), 2.55–2.62 (m, 1H),

2.75–2.79 (m, 2H), 2.80–2.83 (m, 2H), 2.90–2.99 (m, 2H), 3.03– 3.07 (m, 2H), 3.27–3.29 (m, 2H), 3.47–3.53 (m, 2H), 3.58–3.63 (m, 2H), 3.67–3.72 (m, 3H), 3.74–3.79 (m, 2H), 3.80–3.91 (m, 4H), 4.97 (dt, 1H, *J* = 12.5, 3.0 Hz), 6.60 (dd, 1H, *J* = 3.0, 1.5 Hz), 7.07– 7.10 (m, 2H), 7.32 (d, 1H, *J* = 8.5 Hz), 7.45 (d, 1H, *J* = 2.0 Hz), 7.70– 7.71 (m, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 21.0, 21.4, 27.5, 34.9, 36.6, 37.2, 41.5, 42.3, 45.4, 45.9, 46.4, 59.5, 66.7, 109.1, 111.5, 112.0, 117.2, 117.9, 122.3, 125.1, 127.1, 134.0, 137.7, 144.0, 147.5, 159.1, 169.9, 170.4, 172.9; HRMS (ESI) calculated for C₃₂H₃₇N₅O₆Cl 622.2432 [M + H]⁺, found 622.2437.

Indoloquinolizidine 17j. It was prepared from indoloquinolizidine **10m** and amine **16i** in 14.6 mg, 69% yield by following the same protocol of **17a**. Pale yellow liquid: ¹H NMR (500 MHz, CDCl₃) *δ* 1.37 (d, 1H, *J* = 6.5 Hz), 1.68–1.73 (m, 2H), 1.94 (s, 3H), 2.00–2.03 (m, 1H), 2.44–2.51 (m, 2H), 2.69–2.76 (m, 2H), 2.88–2.95 (m, 3H), 3.22–3.29 (m, 3H), 3.30 (s, 3H), 3.31–3.35 (m, 1H), 3.41 (t, 2H, *J* = 5.5 Hz), 3.44–3.48 (m, 1H), 3.49–3.57 (m, 2H), 3.60 (t, 2H, *J* = 5.5 Hz), 3.72–3.76 (m, 1H), 3.84 (s, 3H), 5.06 (dd, 1H, *J* = 12.5, 3.5 Hz), 6.33 (t, 1H, *J* = 5.0 Hz), 6.82 (dd, 1H, *J* = 9.0, 1.5 Hz), 6.92 (d, 1H, *J* = 1.5 Hz), 7.17 (d, 1H, *J* = 9.0 Hz), 8.52 (s, 1H); ¹³C NMR (125 MHz, CDCl₃) *δ* 21.3, 21.8, 26.8, 29.2, 36.0, 37.3, 37.7, 39.8, 42.3, 45.8, 46.4, 55.9, 58.7, 59.7, 66.6, 66.7, 71.2, 100.3, 108.9, 111.8, 112.2, 126.3, 130.7, 137.1, 154.1, 170.5, 171.1, 172.7; HRMS (ESI) calculated for C₂₈H₃₉N₄O₆ 527.2870 [M + H]⁺, found 527.2854.

Hydroxyindoline 11a. A solution of starting indoloquinolizidine 17a (46.3 mg, 0.10 mmol) in CH_2Cl_2 (2.5 mL) was treated with oxaziridine 18 (63.2 mg, 0.32 mmol). The reaction mixture was stirred overnight at room temperature, and then solvent was removed in vacuo. The crude product was purified by flash chromatography on silica gel (elution with EtOAc:MeOH = 4:1) to give hydroxyindoline 11a (25.8 mg, 54%). Pale yellow amorphous solid: ¹H NMR (500 MHz, CD₃OD) δ 1.72 (s, 3H), 1.72–1.75 (m, 1H), 1.87 (ddd, 1H, J = 14.5, 8.0, 3.0 Hz), 2.43-2.47 (m, 1H), 2.48-2.52 (m, 1H), 2.70 (dt, 1H, J = 14.0, 9.0 Hz), 2.83 (s, 3H), 2.83–2.86 (m, 1H), 2.88–2.90 (m, 1H), 2.91 (s, 3H), 3.06–3.11 (m, 1H), 3.99 (dd, 1H, J = 11.0, 3.0 Hz), 4.33 (d, 1H, J = 15.5 Hz), 4.38 (d, 1H, J = 15.5 Hz), 4.62 (ddd, 1H, J = 14.0, 9.0, 3.0 Hz), 6.23 (dd, 1H, J = 3.0, 0.5 Hz), 6.33 (dd, 1H, J = 3.0, 1.5 Hz), 7.27 (dt, 1H, J = 7.5, 1.0 Hz), 7.36 (dt, 1H, J = 7.5, 1.0 Hz), 7.40–7.41 (m, 1H), 7.44 (d, 1H, *J* = 7.0 Hz), 7.46 (d, 1H, *J* = 7.5 Hz); $^{13}\mathrm{C}$ NMR (125 MHz, CD_3OD) δ 20.6, 27.1, 34.9, 36.4, 36.7, 37.2, 37.9, 38.7, 40.0, 40.7, 65.1, 83.1, 108.1, 111.4, 121.8, 123.5, 128.3, 130.5, 143.3, 143.5, 153.0, 154.1, 172.5, 173.6, 187.7; HRMS (ESI) calculated for $C_{26}H_{31}N_4O_5$ 479.2294 [M + H]⁺, found 479.2299. Single crystals for X-ray analysis were obtained by slow evaporation of saturated solution method. CCDC number is 791452.

Hydroxyindoline 11b. A solution of starting indoloquinolizidine 17b (51.5 mg, 0.10 mmol) in CH₂Cl₂ (2.5 mL) was treated with oxaziridine 18 (63.2 mg, 0.32 mmol). The reaction mixture was stirred overnight at room temperature, and then solvent was removed in vacuo. The crude product was purified by flash chromatography on silica gel (elution with EtOAc:MeOH = 4:1) to give hydroxyindoline 11b (34.5 mg, 65%). Pale yellow amorphous solid: ¹H NMR (500 MHz, CD₃OD) δ 1.32 (s, 9H), 1.70–1.72 (m, 1H), 1.73 (s, 3H), 1.94 (ddd, 1H, J = 14.5, 8.0, 3.0 Hz), 2.42 (dd, 1H, J = 14.5, 10.0 Hz),2.51-2.57 (m, 1H), 2.65-2.70 (m, 1H), 2.73 (dd, 1H, J = 14.5, 4.0 Hz), 2.89 (dt, 1H, J = 14.0, 8.0 Hz), 3.01-3.05 (m, 1H), 3.11-3.15 (m, 1H), 3.33–3.36 (m, 2H), 3.40–3.56 (m, 3H), 3.61–3.69 (m, 2H), 3.95 (dd, 1H, J = 11.5, 3.5 Hz), 4.59 (ddd, 1H, J = 14.0, 9.0, 3.0 Hz), 7.40 (dd, 1H, J = 8.0, 2.0 Hz), 7.45 (d, 1H, J = 2.0 Hz), 7.47 (d, 1H, J = 8.0 Hz), 7.67 (s, 1H); ¹³C NMR (125 MHz, CD₃OD) δ 20.1, 27.3, 28.9, 35.2, 36.8, 37.9, 40.1, 40.9, 43.7, 48.0, 52.2, 65.2, 67.5, 67.7, 83.3, 123.1, 124.2, 130.5, 134.1, 145.7, 152.6, 171.2, 173.1, 173.7, 187.9; HRMS (ESI) calculated for $C_{27}H_{36}N_4O_5Cl 531.2374 [M + H]^+$, found 531.2366.

Hydroxyindoline 11c. A solution of starting indoloquinolizidine 17c (43.9 mg, 0.10 mmol) in CH_2Cl_2 (2.5 mL) was treated with oxaziridine 18 (63.2 mg, 0.32 mmol). The reaction mixture was stirred overnight at room temperature, and then solvent was removed in vacuo. The crude product was purified by flash chromatography on silica gel (elution with EtOAc:MeOH = 4:1) to give hydroxyindoline

11c (31.4 mg, 69%). Pale yellow amorphous solid: ¹H NMR (500 MHz, CD₃OD) δ 1.32 (s, 9H), 1.72 (s, 3H), 1.72–1.73 (m, 1H), 1.85–1.90 (m, 1H), 2.39 (ddd, 1H, *J* = 14.5, 10.0, 2.0 Hz), 2.45–2.52 (m, 1H), 2.67–2.71 (m, 1H), 2.75 (ddd, 1H, *J* = 14.5, 4.0, 1.5 Hz), 2.84 (s, 3H), 2.86–2.90 (m, 1H), 2.92 (s, 3H), 3.03–3.05 (m, 1H), 3.99–4.02 (m, 1H), 4.59–4.64 (m, 1H), 7.26–7.29 (m, 1H), 7.35–7.38 (m, 1H), 7.44–7.47 (m, 2H), 7.68 (s, 1H); ¹³C NMR (125 MHz, CD₃OD) δ 20.5, 27.2, 28.9, 35.1, 36.4, 36.7, 38.0, 38.7, 40.7, 41.0, 52.0, 65.1, 83.1, 121.8, 123.5, 128.3, 130.5, 143.5, 154.1, 172.5, 173.2, 173.9, 187.7; HRMS (ESI) calculated for C₂₅H₃₅N₄O₄ 455.2658 [M + H]⁺, found 455.2647.

Spirocyclic Dihydroindolone 12a. A solution of starting hydroxyindoline 11b (5.10 mg, 9.6 µmol) in MeOH (0.6 mL) was treated with aqueous H_2SO_4 (0.2 mL, 10% v/v). The reaction mixture was stirred overnight at 40 °C and then diluted to 1.0 mL. The crude product was purified by LC-MS to give spirocyclic dihydroindolone 12a (4.69 mg, 92%). Pale yellow amorphous solid: ¹H NMR (500 MHz, CD_3OD) δ 1.32 (s, 9H), 1.61 (d, 1H, J = 14.0 Hz), 1.72 (s, 3H), 2.01 (dd, 1H, J = 13.5, 8.5 Hz), 2.27-2.37 (m, 2H), 2.40 (dd, 1H, J = 15.0, 9.0 Hz), 2.64 (dd, 2H, J = 15.0, 4.0 Hz), 2.90-2.97 (m, 1H), 3.01-3.03 (m, 1H), 3.15 (dd, 1H, J = 13.0, 4.0 Hz), 3.20-3.23 (m, 1H), 3.34-3.39 (m, 5H), 3.61 (t, 1H, J = 11.0 Hz), 3.79-3.85 (m, 1H), 6.87 (d, 1H, J = 8.5 Hz), 7.37 (s, 1H), 7.42 (d, 1H, J = 9.0 Hz), 7.47 (s, 1H), 8.16 (s, 1H); ¹³C NMR (125 MHz, CD₃OD) δ 20.5, 28.2, 29.0, 29.8, 37.3, 38.6, 40.4, 42.4, 43.0, 47.7, 52.1, 67.2, 67.3, 67.6, 77.7, 115.0, 121.1, 124.1, 124.3, 138.9, 161.4, 171.7, 172.7, 172.8, 203.5; HRMS (ESI) calculated for C₂₇H₃₆N₄O₅Cl 531.2374 [M + H]⁺, found 531.2377. Single crystals for X-ray analysis were obtained by slow evaporation of saturated solution method. CCDC number is 791453.

Spirocyclic Dihydroindolone 12b. A solution of starting hydroxyindoline 11c (4.38 mg, 9.6 μ mol) in MeOH (0.6 mL) was treated with aqueous H_2SO_4 (0.2 mL, 10% v/v). The reaction mixture was stirred overnight at 40 °C and then diluted to 1.0 mL. The crude product was purified by LC-MS to give spirocyclic dihydroindolone 12b (2.85 mg, 65%). Pale yellow amorphous solid: ¹H NMR (500 MHz, CDCl₃) δ 1.36 (s, 9H), 1.78 (s, 3H), 2.00–2.04 (m, 1H), 2.07 (ddd, 1H, J = 13.0, 8.0, 1.5 Hz), 2.20 (s, 3H), 2.22-2.30 (m, 2H), 2.36 (dd, 1H, J = 13.5, 5.0 Hz), 2.66 (s, 3H), 2.71 (dd, 1H, J = 13.5, 6.0 Hz), 2.86–2.88 (m, 1H), 2.93 (dd, 1H, J = 13.5, 5.0 Hz), 3.68 (t, 1H, J = 11.0 Hz), 3.94-4.00 (m, 1H), 4.78 (s, 1H), 6.26 (s, 1H), 6.82 (dt, 1H, J = 8.0, 0.5 Hz), 6.89 (d, 1H, J = 8.5 Hz), 7.41–7.44 (m, 1H), 7.55-7.57 (m, 1H); ¹³C NMR (125 MHz, CD₃OD) δ 20.5, 27.9, 29.0, 29.6, 35.6, 37.4, 38.0, 38.8, 40.8, 42.5, 52.1, 67.3, 76.8, 113.4, 119.2, 120.3, 124.9, 139.2, 163.1, 172.7, 172.9, 173.0, 205.1; HRMS (ESI) calculated for C₂₅H₃₅N₄O₄ 455.2658 [M + H]⁺, found 455.2665.

Tricylic Ketoamide 13a. A solution of starting indologuinolizidine 17b (61.4 mg, 0.14 mmol) in CH_2Cl_2 (3 mL) was treated with m-CPBA (95.0 mg, 0.42 mmol). The reaction mixture was stirred for 30 min at room temperature, diluted with EtOAc (6 mL), washed with Na₂SO₃ (saturated solution, 5 mL) and brine (5 mL), dried over MgSO₄, filtered and concentrated in vacuo. The crude product was purified by flash chromatography on silica gel (elution with EtOAc:MeOH = 4:1) to give tricyclic ketoamide 13a (46.8 mg, 71%). Pale yellow amorphous solid: ¹H NMR (500 MHz, CDCl₃) δ 1.26 (s, 9H), 1.44 (s, 3H), 2.12 (dt, 1H, J = 14.5, 4.5 Hz), 2.19 (dd, 1H, J = 16.0, 3.0 Hz), 2.33–2.40 (m, 1H), 2.82–2.87 (m, 1H), 2.94– 2.98 (m, 4H), 3.03 (dd, 1H, J = 13.5, 6.5 Hz), 3.09 (dd, 1H, J = 14.5, 6.5 Hz), 3.17 (s, 3H), 3.25 (dd, 1H, J = 16.0, 6.0 Hz), 3.84 (dd, 1H, J = 9.0, 4.0 Hz), 4.54 (dd, 1H, J = 15.0, 11.0 Hz), 5.59 (s, 1H), 7.17 (d, 1H, J = 8.0 Hz), 7.41 (t, 1H, J = 7.5 Hz), 7.54 (dt, 1H, J = 7.5, 1.5 Hz), 7.61 (dd, 1H, J = 7.5, 1.5 Hz), 10.64 (s, 1H); ¹³C NMR (125 MHz, $CDCl_3$) δ 19.3, 27.6, 28.7, 34.7, 36.2, 37.6, 38.9, 39.9, 40.7, 43.9, 52.0, 67.0, 127.8, 127.9, 128.8, 132.2, 136.7, 140.4, 170.5, 170.5, 171.9, 174.6, 204.5; HRMS (ESI) calculated for C₂₅H₃₅N₄O₅ 471.2607 [M + H]⁺, found 471.2598. Single crystals for X-ray analysis were obtained by slow evaporation of saturated solution method. CCDC numer is 791450.

Tricyclic Ketoamide 13c. It was prepared in 16.9 mg, 32% yield by following the same procotol of 13a. Pale yellow amorphous solid:

¹H NMR (500 MHz, CDCl₃) δ 1.45 (s, 3H), 2.22–2.27 (m, 1H), 2.27–2.31 (m, 2H), 2.81–2.83 (m, 1H), 2.90–2.94 (m, 1H), 2.95 (s, 3H), 3.03–3.05 (m, 1H), 3.06–3.08 (m, 1H), 3.13 (s, 3H), 3.32 (dd, 1H, *J* = 16.0, 6.0 Hz), 3.78 (dd, 1H, *J* = 7.5, 4.5 Hz), 4.20 (dd, 1H, *J* = 15.5, 5.0 Hz), 4.39 (dd, 1H, *J* = 15.5, 6.5 Hz), 4.54 (dd, 1H, *J* = 14.0, 4.0 Hz), 6.09–6.10 (m, 1H), 6.25–6.28 (m, 2H), 7.17 (d, 1H, *J* = 8.5 Hz), 7.30–7.31 (m, 1H), 7.50 (dd, 1H, *J* = 8.5, 2.5 Hz), 7.58 (d, 1H, *J* = 2.5 Hz), 10.4 (s, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 19.7, 27.3, 34.4, 36.2, 36.7, 37.7, 38.5, 38.8, 40.3, 43.8, 67.1, 107.8, 110.5, 128.9, 129.7, 132.1, 134.0, 135.2, 141.5, 142.3, 150.4, 170.4, 170.7, 172.3, 174.8, 203.0; HRMS (ESI) calculated for C₂₆H₃₀N₄O₆Cl 529.1854 [M + H]⁺, found 529.1861.

Pvrroloquinolone 14a. A solution of starting tricyclic ketoamide 13b (33.8 mg, 0.062 mmol) in EtOH (1 mL) was treated with KOH (4.3 mg, 0.077 mmol). The reaction mixture was stirred overnight at room temperature, and then solvent was removed in vacuo. The crude product was purified by flash chromatography on silica gel (elution with EtOAc:MeOH = 4:1) to give pyrroloquinolone 14a (17.3 mg, 53%). Pale yellow amorphous solid: ¹H NMR (500 MHz, CDCl₃) δ 1.27 (s, 9H), 1.55-1.61 (m, 5H), 1.92 (s, 3H), 2.26-2.31 (m, 1H), 2.34 (dd, 1H, J = 14.5, 5.0 Hz), 2.38-2.46 (m, 1H), 2.89-2.92 (m, 1H), 2.97 (dd, 1H, J = 14.5, 4.5 Hz), 3.26 (dd, 1H, J = 13.5, 6.0 Hz), 3.39-3.48 (m, 3H), 3.59-3.62 (m, 1H), 3.74-3.77 (m, 1H), 4.55 (d, 1H, J = 15.0 Hz), 4.92 (d, 1H, J = 15.5 Hz), 5.95 (s, 1H), 7.28 (d, 1H, J = 9.0 Hz, 7.51 (dd, 1H, J = 9.0, 2.5 Hz), 8.33 (d, 1H, J = 2.5 Hz), 10.3 (s, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 22.6, 24.2, 25.8, 26.7, 26.9, 28.7, 35.5, 39.0, 43.5, 47.2, 47.3, 48.2, 51.1, 67.9, 113.3, 119.7, 125.3, 126.8, 129.7, 132.1, 138.1, 152.6, 169.2, 170.5, 171.3, 173.5; HRMS (ESI) calculated for $C_{28}H_{36}N_4O_4Cl 527.2425 [M + H]^+$, found 527.2418. Single crystals for X-ray analysis were obtained by slow evaporation of saturated solution method. CCDC number is 791454.

Pyrroloquinolone 14b. It was prepared in 18.3 mg, 69% yield by following the same procotol of **14a**. Pale yellow amorphous solid: ¹H NMR (500 MHz, CDCl₃) δ 1.26 (s, 9H), 1.90 (s, 3H), 2.26 (dt, 1H, *J* = 12.5, 5.0 Hz), 2.33 (dd, 1H, *J* = 14.5, 4.5 Hz), 2.41–2.48 (m, 1H), 2.86–2.89 (m, 1H), 3.01 (dd, 1H, *J* = 15.0, 4.5 Hz), 3.35 (dd, 1H, *J* = 13.5, 6.0 Hz), 3.56–3.60 (m, 2H), 3.64–3.66 (m, 2H), 3.70–3.73 (m, 2H), 3.75–3.79 (m, 2H), 4.54 (d, 1H, *J* = 15.0 Hz), 4.90 (d, 1H, *J* = 15.0 Hz), 5.88 (s, 1H), 7.28 (s, 1H), 7.49 (dd, 1H, *J* = 9.0, 2.5 Hz), 8.29 (d, 1H, *J* = 2.5 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 22.7, 27.1, 28.7, 35.3, 38.9, 42.6, 46.7, 46.7, 48.2, 51.1, 66.7, 66.8, 67.7, 113.3, 119.8, 125.2, 126.7, 129.7, 132.1, 138.1, 152.4, 169.0, 170.6, 172.0, 173.4; HRMS (ESI) calculated for C₂₇H₃₄N₄O₅Cl 529.2218 [M + H]⁺, found 529.2205.

Syntheses of 476-Member Library of Indoloquinolizidines 17. Forty quinolizidine-containing carboxylic acids **10** were coupled with 12 amines **16** (1.5 equiv) and then treated with HATU (1.5 equiv), and DIPEA (3 equiv) in DMF. The reaction mixtures were stirred overnight at room temperature. The 480 crude mixtures were purified by preparative LC–MS to give 476 indoloquinolizidines **17** with an average yield of 20 mg (65%) and greater than 95% purity.

Syntheses of 96-Member Library of Indoloquinolizidines 17. Eight quinolizidine-containing carboxylic acids 10 were coupled with 12 amines 16 (1.5 equiv) and then treated with HATU (1.5 equiv), and DIPEA (3 equiv) in DMF. The reaction mixtures were stirred overnight at room temperature. The 96 crude mixtures were purified by preparative LC–MS to give 96 indoloquinolizidines 17 with an average yield of 58 mg (74%) and greater than 95% purity. The products were then split into two sets of vials for different diversification pathways.

Syntheses of 83-Member Library of Spirocyclic Dihydroindolones 12. Ninety-six indoloquinolizidines 17 were treated with oxaziridine 18 (3 equiv) in CH_2Cl_2 . The reaction mixtures were stirred overnight at room temperature. The 96 crude mixtures were purified by preparative LC–MS to give 96 hydroxyindolines 11, which were immediately dissolved in MeOH and treated with aqueous H_2SO_4 (10% v/v). The reaction mixtures were stirred overnight at 40 °C and then diluted by water. The 96 crude mixtures were purified by preparative LC–MS to give 83 spirocyclic dihydroindolones 12 with

an average yield of 7 mg (30%) after two steps and greater than 95% purity.

Syntheses of 96-Member Library of Tricyclic Ketoamides 13. Ninety-six indoloquinolizidines 17 were treated with *m*-CPBA (3 equiv) in CH_2Cl_2 . The reaction mixtures were stirred for 30 min at room temperature, diluted with EtOAc, and washed with Na_2SO_3 (saturated solution) and brine. The 96 crude mixtures were purified by preparative LC-MS to give 96 tricyclic ketoamide 13 with an average yield of 11 mg (47%) and greater than 95% purity. The products were then split into two sets of vials for final diversification.

Syntheses of 96-Member Library of Pyrroloquinolones 14. Ninety-six tricyclic ketoamides 13 in EtOH were treated with KOH (1.0 equiv). The reaction mixtures were stirred overnight at room temperature. The 96 crude mixtures were purified by preparative LC–MS to give 96 pyrroloquinolones 14 with an average yield of 5 mg (64%) and greater than 95% purity.

ASSOCIATED CONTENT

S Supporting Information

Detailed experimental procedures, crystal data (CIF), and analytical and spectral characterization data for all new compounds. This information is available free of charge via the Internet at http://pubs.acs.org.

AUTHOR INFORMATION

Corresponding Author

*E-mail: skozmin@uchicago.edu.

Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

This work was funded by the National Institutes of Health (P50 GM086145) and the Chicago Biomedical Consortium with support from the Searle Funds at the Chicago Community Trust. We thank Dr. Ian Steele for X-ray crystallographic analysis.

REFERENCES

(1) (a) Cane, D. E.; Walsh, C. T.; Khosla, C. Science **1998**, 282, 63– 68. (b) Walsh, C. ChemBioChem **2002**, 3, 124–134. (c) Weissman, K. J.; Leadlay, P. F. Nat. Rev. Microbiol. **2005**, 3, 925–936.

(2) (a) Hall, D. G.; Manku, S.; Wang, F. J. Comb. Chem. 2001, 3, 125–150.
(b) Breinbauer, M.; Manger, M.; Scheck, H.; Waldmann,. Curr. Med. Chem. 2002, 9, 2129–2145.
(c) Nielsen, J. Curr. Opin. Chem. Biol. 2002, 6, 297–305.
(d) Boldi, A. M. Curr. Opin. Chem. Biol. 2004, 8, 281–286.
(e) Nandy, J. P.; Prakesch, M.; Khadem, S.; Reddy, P. T.; Sharma, U.; Arya, P. Chem. Rev. 2009, 109, 1999–2060.

(3) (a) Pelish, H. E.; Westwood, N. J.; Feng, Y.; Kirchhausen, T.; Shair, M. D. J. Am. Chem. Soc. 2001, 123, 6740–6741. (b) Goess, B.; Hannoush, R.; Chan, L.; Kirchhausen, T.; Shair, M. D. J. Am. Chem. Soc. 2006, 128, 5391–5403.

- (4) O'Connor, S. E.; Maresh, J. J. Nat. Prod. Rep. 2006, 23, 532-547.
- (5) Treimer, J. F.; Zenk, M. H. Eur. J. Biochem. 1979, 101, 225-233.
- (6) Scott, A. I. Acc. Chem. Res. 1970, 3, 151–157.

(7) Hutchinson, C. R.; Heckendorf, A. H.; Daddona, P. E.; Hagaman, E.; Wenkert., E. J. Am. Chem. Soc. **1974**, *96*, 5609–5611.

(8) McCoy, E.; O'Connor, S. E. J. Am. Chem. Soc. 2006, 128, 14276–14277.

(9) (a) Bernhardt, P.; McCoy, E.; O'Connor, S. E. *Chem. Biol.* 2007, *14*, 888–897. (b) Loris, A.; Panjikar, S.; Ruppert, M.; Barleben, L.; Unger, M.; Schübel, H.; Stöckigt, J. *Chem. Biol.* 2007, *14*, 979–985. (c) Runguphan, W.; O'Connor, S. E. *Nat. Chem. Biol.* 2009, *5*, 151–153.

(10) (a) Abelman, M. M.; Curtis, J. K.; James, D. R. *Tetrahedron Lett.* **2003**, 44, 6527–6531. (b) Karpov, A. S.; Oeser, T.; Müller, T. J. J. *Chem. Commun.* **2004**, 1502–1503. (11) Davis, F. A.; Towson, J. C.; Vashi, D. B.; ThimmaReddy, R.; McCauley, J. P.; Harakal, M. E.; Gosciniak, D. J. *J. Org. Chem.* **1990**, 55, 1254–1261.

(12) (a) Stahl, R.; Borschberg, H. J. Helv. Chim. Acta 1994, 77, 1331–1345. (b) Güller, R.; Borschberg, H.-J. Tetrahedron: Asymmetry 1992, 3, 1197–1204.

(13) Lemaire, S.; Willemsens, B.; Marko, I. E. Synlett 2007, 709–712.
(14) Details of the screening results are deposited in PubChem: http://pubchem.ncbi.nlm.nih.gov.

(15) Martinez, A.; Ulanovskaya, O. A.; Zhang, R.; Cui, J.; Schumacker, P. T.; Kozmin, S. A. unpublished results.

(16) For representative examples, see: (a) Ding, S.; Gray, N. S.; Wu, X.; Ding, Q.; Schultz, P. G. J. Am. Chem. Soc. 2002, 124, 1594–1595.
(b) Kwon, O.; Park, S. B.; Schreiber, S. L. J. Am. Chem. Soc. 2002, 124, 13402–13404. (c) Burke, M. D.; Berger, E. M.; Schreiber, S. L. Science 2003, 302, 613–618. (d) Oguri, H.; Schreiber, S. L. Org. Lett. 2005, 7, 47–50. (e) Spiegel, D. A.; Schroeder, F. C.; Duvall, J. R.; Schreiber, S. L. J. Am. Chem. Soc. 2007, 129, 1020–1021. (g) Morton, D.; Leach, S.; Cordier, C.; Warriner, S.; Nelson M., A. Angew. Chem., Int. Ed. 2009, 48, 104–109. (h) Medeiros, R.; Narayan, R. S.; McDougal, N. T.; Schaus, S. E.; Porco, J. A. Org. Lett. 2010, 12, 3222–3225. (i) Thomas, G. L.; Spandl, R. J.; Glansdorp, F. G.; Welch, M.; Bender, A.; Cockfield, J.; Lindsay, J. A.; Bryant, C.; Brown, D. F. J.; Loiseleur, O.; Rudyk, H.; Ladlow, M.; Spring, D. R. Angew. Chem., Int. Ed. 2008, 47, 2808–2812.

(17) Clardy, J.; Walsh, C. Nature 2004, 432, 830-837.